Structured Search

Database: US Pre-Grant Publication Full-Text Database; US Patents Full-Text Database; US OCR Full-Text Database; EPO Abstracts Database; JPO Abstracts Database; Derwent World Patents Index; IBM Technical Disclosure Bulletins

Searchable Index: <none> - All Fields | Not Numeric

Term 1 text:

Operator: AND **Proximity Distance:** 1

Searchable Index: <none> - All Fields | Not Numeric

Term 2 text:

Display: 10 **Documents in Display Format:** TI,KWIC **Starting With #:** 1

Generate: ○ Hit List ◉ Hit Count ○ Side by Side ○ Image

Search Clear Interrupt

Search History

DATE: Wednesday, June 07, 2006 Printable Copy Create Case

Set Name side by side	Query	Hit Count	Set Name result set
DB=USPT; PLUR=YES; OP=ADJ			
L13	(l8 near15 l9 near15 blood)	24	L13
L12	l8 near30 l9 near30 l10	8	L12
L11	l8 near12 l9 near12 l10	1	L11
L10	platelet	37402	L10
L9	phosphate or PO4	251698	L9
L8	L7 or disaccharide	118915	L8
L7	trehalose or maltose or cellobiose or lactose or sucrose	116848	L7
L6	(l1 adj2 esterified)[CLM]	107	L6
L5	fatty acid is esterified[clm]	0	L5
L4	(wherein adj2 l1)	0	L4
L3	l1 adj4 (ester form or esterified)	1813	L3
L2	L1 adj3 in the form of adj4 ester	0	L2

Hit List

First Hit | Clear | Generate Collection | Print | Fwd Refs | Bkwd Refs | Generate OACS

Search Results - Record(s) 1 through 8 of 8 returned.

☐ 1. Document ID: US 6632457 B1

L12: Entry 1 of 8 File: USPT Oct 14, 2003

US-PAT-NO: 6632457
DOCUMENT-IDENTIFIER: US 6632457 B1
** See image for Certificate of Correction **

TITLE: Composite hydrogel drug delivery systems

L12: Entry 1 of 8 File: USPT Oct 14, 2003

DOCUMENT-IDENTIFIER: US 6632457 B1
** See image for Certificate of Correction **
TITLE: Composite hydrogel drug delivery systems

Detailed Description Text (64):
The preferred binding ligand, heparin, has been shown to form affinity-bound complexes with a number of active agents, including without limitation: antithrombin III; Factors VII, IX, XI, XII, and XIIa; thrombin; properdin; complements C1, C2, C3 and C4; complement factor .beta.; C3b inactivator; Gc globulin; protein HC; fibronectin; .beta.2-glycoprotein 1; C-reactive protein; lipoprotein lipase; hepatic triglyceride lipase; VLDL, LDL; VLDL apoprotein; HDLP; restriction endonucleases; RNA polymerase; RNA polymerases I and II; DNA polymerase; DNA ligase; polynucleotide kinase; elongation factor (EF-1); initiation factors; protein synthesis factors; ribosomes; estrogen receptor; androgen receptor; platelet factor 4; SV 40 tumor antigen; Hepatitis B surface antigen; hyaluronidase; collagenase inhibitor; neurophysin; and trehalose phosphate synthetase.

Full | Title | Citation | Front | Review | Classification | Date | Reference | Sequences | Attachments | Claims | KWIC | Draw D

☐ 2. Document ID: US 6610291 B2

L12: Entry 2 of 8 File: USPT Aug 26, 2003

US-PAT-NO: 6610291
DOCUMENT-IDENTIFIER: US 6610291 B2

TITLE: Ready-to-use ristocetin cofactor test reagent possessing long-term stability

L12: Entry 2 of 8 File: USPT Aug 26, 2003

DOCUMENT-IDENTIFIER: US 6610291 B2
TITLE: Ready-to-use ristocetin cofactor test reagent possessing long-term stability

Detailed Description Text (5):
The platelet suspension was then dialyzed against the 20-fold volume of phosphate buffer C for a period of approx. 24 hours. After that, the platelet count was adjusted by adding phosphate buffer C. The following were also added: sucrose (10 g/l), glycine (10 g/l), glutamate (16.7 g/l) and human albumin (2 g/l).

Full | Title | Citation | Front | Review | Classification | Date | Reference | Sequences | Attachments | Claims | KWIC | Draw D

☐ 3. Document ID: US 6518244 B2

L12: Entry 3 of 8 File: USPT Feb 11, 2003

US-PAT-NO: 6518244
DOCUMENT-IDENTIFIER: US 6518244 B2
**** See image for Certificate of Correction ****

TITLE: Combinations of heparin cofactor II agonist and platelet IIb/IIIa antagonist, and uses thereof

L12: Entry 3 of 8 File: USPT Feb 11, 2003

DOCUMENT-IDENTIFIER: US 6518244 B2
**** See image for Certificate of Correction ****
TITLE: Combinations of heparin cofactor II agonist and platelet IIb/IIIa antagonist, and uses thereof

Detailed Description Text (20):
HCII agonists and/or platelet GPIIb/IIIa receptor antagonists that are orally active can be administered as oral dose forms one or more times during the day, e.g., one, two, three or four times daily. For oral administration in the form of a tablet or capsule, the active ingredient (i.e., the HCII agonists, the platelet GPIIb/IIIa receptor antagonist or both) can be combined with an oral, non-toxic pharmaceutically acceptable, inert carrier such as lactose, starch, sucrose, glucose, methyl cellulose, magnesium stearate, dicalcium phosphate, calcium sulfate, mannitol, sorbitol and the like. For oral administration in liquid form, the oral drug components can be combined with any oral, non-toxic, pharmaceutically acceptable inert carrier such as ethanol, glycerol, water and the like. Moreover, when desired or necessary, suitable binders, lubricants, disintegrating agents and coloring agents can also be incorporated into the mixture. Suitable binders include starch, gelatin, natural sugars such as glucose or .beta.-lactose, corn-sweeteners, natural and synthetic gums such as acacia, tranacanth or sodium alginate, carboxymethylcellulose, polyethylene glycol, waxes and the like. Lubricants used in these dosage forms include sodium oleate, sodium stearate, magnesium stearate, sodium benzoate, sodium acetate, sodium chloride and the like. Disintegrators include, without limitation, starch, methyl cellulose, agar, bentonite, xanthan gum and the like.

Full | Title | Citation | Front | Review | Classification | Date | Reference | Sequences | Attachments | Claims | KWIC | Draw D

☐ 4. Document ID: US 6007978 A

L12: Entry 4 of 8 File: USPT Dec 28, 1999

US-PAT-NO: 6007978
DOCUMENT-IDENTIFIER: US 6007978 A

TITLE: Method of freezing cells and cell-like materials

L12: Entry 4 of 8 File: USPT Dec 28, 1999

DOCUMENT-IDENTIFIER: US 6007978 A
TITLE: Method of freezing cells and cell-like materials

Detailed Description Text (113):
To extend the study in Example 9, we again prepared fresh, one day-old human platelet concentrates mixed 1:1 by volume with the lactose/mannose/PVP-10 formulation (LMP) as described in Example 1 (the solution is formulated as 10% w/v lactose, 5% w/v mannose, 10% w/v PVP-10,000 mM in 10 MW phosphate buffer, pH 7.2, and is diluted into an equal volume of platelet concentrate). We then compared the ability of this formulation to preserve human platelets at elevated frozen storage temperatures as high as -25.degree. C., relative to standard 5% DMSO. In this experiment four independent samples were prepared and frozen for each parameter (fresh controls, DMSO, and LMP), and then stored at three different temperatures: -80.degree. C., -38.degree. C., and -25.degree. C. In Table 14 we show the data obtained after the frozen platelets were stored for 7 days at each temperature, and then thawed and prepared for aggregometry analysis by a single centrifugation step to remove most of the cryoprotectants. In Table 4 the aggregation response values are expressed as chart percentages. Morphology score and percent cell recovery are obtained as described in Example 7.

☐ 5. Document ID: US 5958670 A

L12: Entry 5 of 8 File: USPT Sep 28, 1999

US-PAT-NO: 5958670
DOCUMENT-IDENTIFIER: US 5958670 A
** See image for Certificate of Correction **

TITLE: Method of freezing cells and cell-like materials

L12: Entry 5 of 8 File: USPT Sep 28, 1999

DOCUMENT-IDENTIFIER: US 5958670 A
** See image for Certificate of Correction **
TITLE: Method of freezing cells and cell-like materials

Detailed Description Text (112):
To extend the study in Example 9, we again prepared fresh, one day-old human

platelet concentrates mixed 1:1 by volume with the lactose/mannose/PVP-10 formulation (LMP) as described in Example 1 (the solution is formulated as 10% w/v lactose, 5% w/v mannose, 10% w/v PVP-10,000 mM in 10 MW phosphate buffer, pH 7.2, and is diluted into an equal volume of platelet concentrate). We then compared the ability of this formulation to preserve human platelets at elevated frozen storage temperatures as high as -25.degree. C., relative to standard 5% DMSO. In this experiment four independent samples were prepared and frozen for each parameter (fresh controls, DMSO, and LMP), and then stored at three different temperatures: -80.degree. C., -38.degree. C., and -25.degree. C. In Table 14 we show the data obtained after the frozen platelets were stored for 7 days at each temperature, and then thawed and prepared for aggregometry analysis by a single centrifugation step to remove most of the cryoprotectants. In Table 4 the aggregation response values are expressed as chart percentages. Morphology score and percent cell recovery are obtained as described in Example 7.

☐ 6. Document ID: US 5800978 A

L12: Entry 6 of 8 File: USPT Sep 1, 1998

US-PAT-NO: 5800978
DOCUMENT-IDENTIFIER: US 5800978 A
**** See image for Certificate of Correction ****

TITLE: Method of freezing cells and cell-like materials

L12: Entry 6 of 8 File: USPT Sep 1, 1998

DOCUMENT-IDENTIFIER: US 5800978 A
**** See image for Certificate of Correction ****
TITLE: Method of freezing cells and cell-like materials

Detailed Description Text (110):
To extend the study in Example 9, we again prepared fresh, one day-old human platelet concentrates mixed 1:1 by volume with the lactose/mannose/PVP-10 formulation (LMP) as described in Example 1 (the solution is formulated as 10% w/v lactose, 5% w/v mannose, 10% w/v PVP-10,000 mM in 10 MW phosphate buffer, pH 7.2, and is diluted into an equal volume of platelet concentrate). We then compared the ability of this formulation to preserve human platelets at elevated frozen storage temperatures as high as -25.degree. C., relative to standard 5% DMSO. In this experiment four independent samples were prepared and frozen for each parameter (fresh controls, DMSO, and LMP), and then stored at three different temperatures: -80.degree. C., -38.degree. C., and -25.degree. C. In Table 14 we show the data obtained after the frozen platelets were stored for 7 days at each temperature, and then thawed and prepared for aggregometry analysis by a single centrifugation step to remove most of the cryoprotectants. In Table 4 the aggregation response values are expressed as chart percentages. Morphology score and percent cell recovery are obtained as described in Example 7.

Full | Title | Citation | Front | Review | Classification | Date | Reference | Sequences | Attachments | Claims | KWIC | Draw D

☐ 7. Document ID: US 5693341 A

L12: Entry 7 of 8 File: USPT Dec 2, 1997

US-PAT-NO: 5693341
DOCUMENT-IDENTIFIER: US 5693341 A

TITLE: Affinity bound collagen matrices for the delivery of biologically active agents

L12: Entry 7 of 8 File: USPT Dec 2, 1997

DOCUMENT-IDENTIFIER: US 5693341 A
TITLE: Affinity bound collagen matrices for the delivery of biologically active agents

Detailed Description Text (11):
The preferred binding ligand, heparin, has been shown to form affinity bound complexes with a number of active agents, including without limitation: antithrombin III; Factors VII, IX, XI, XII, and XIIa; thrombin; properdin; complements C1, C2, C3, and C4; complement factor B; C3b inactivator; Gc globulin; protein HC; fibronectin; .beta.2-glycoprotein 1; C-reactive protein; lipoprotein lipase; hepatic triglyceride lipase; VLDL, LDL; VLDL apoprotein; HDLP; restriction endonucleases; RNA polymerase; RNA polymerase I and II; DNA polymerase; DNA ligase; polynucleotide kinase; elongation factor (EF-1); initiation factors; protein synthesis factors; ribosomes; estrogen receptor; androgen receptor; platelet factor 4; SV 40 tumor antigen; Hepatitis B surface antigen; hyaluronidase; collagenase inhibitor; neurophysin; trehalose phosphate synthetase. Heparin is also known to form affinity bound complexes with the following agents: transforming growth factor beta (TGF-.beta.), fibroblast growth factor (FGF), platelet-derived growth factor (PDGF), epidermal growth factor (EGF), osteogenin, insulin-like growth factors (IGFs), vascular endothelial growth factor, granulocyte/macrophage colony-stimulating factor (CSF), gamma interferon, glia-activating factors, and collagen type V.

☐ 8. Document ID: US 5569579 A

L12: Entry 8 of 8 File: USPT Oct 29, 1996

US-PAT-NO: 5569579
DOCUMENT-IDENTIFIER: US 5569579 A

TITLE: Synthetic-based platelet storage media

L12: Entry 8 of 8 File: USPT Oct 29, 1996

DOCUMENT-IDENTIFIER: US 5569579 A
TITLE: Synthetic-based platelet storage media

Brief Summary Text (17):

Shimizu et al., "Plasma-poor Platelet Concentrates (PC) Prepared by Autoclave-Sterilized Additive Solution Containing Glucose With Physiological pH" (1990) sets forth a synthetic storage medium containing added glucose, maltose, phosphate, and acetate wherein the amount of phosphate and acetate is above 20 mM.

Term	Documents
(10 NEAR30 (8 NEAR30 9)).USPT.	8
(L8 NEAR30 L9 NEAR30 L10).USPT.	8

Hit List

First Hit | Clear | Generate Collection | Print | Fwd Refs | Bkwd Refs | Generate OACS

Search Results - Record(s) 1 through 10 of 24 returned.

☐ 1. Document ID: US 7029564 B1

L13: Entry 1 of 24 File: USPT Apr 18, 2006

US-PAT-NO: 7029564
DOCUMENT-IDENTIFIER: US 7029564 B1

TITLE: Dielectrophoretic apparatus and method

L13: Entry 1 of 24 File: USPT Apr 18, 2006

DOCUMENT-IDENTIFIER: US 7029564 B1
TITLE: Dielectrophoretic apparatus and method

Description Paragraph (42):
In one experiment, a 6 milliliter sample of human whole blood was collected in a lithium heparin tube, and within one hour was diluted 40 times in a phosphate-buffered saline solution containing sucrose, glucose, heparin and calcium chloride, to give a final suspension conductivity of 15 mS/m. The serpentine electrodes were energised with a 20 kHz, 0.6 Vrms stationary DEP signal so as to levitate the blood cells above the electrode plane when they were introduced into the test chamber. This DEP signal was then removed and two TWD signals were applied to the electrodes, one comprising a 50 kHz, 0.32 Vrms forward travelling wave and the other a 400 kHz, 0.64 Vrms reverse travelling wave. The majority of the blood cells moved rapidly along channels 3 and 5 similar to the case shown in FIG. 4f, principally under the action of the 50 kHz signal. A small number, of the order 5% or less of the total number, of the blood cells were found to be trapped on the electrodes or to move slowly along channels 2 and 4 similar to the case shown in FIG. 4a. Microscopic inspection, using a .times.40 objective, indicated that approximately 20 25 red blood cells were trapped or moving in channels 2 and 4 for every white blood cell. On re-applying the 20 kHz stationary DEP signal, the trapped red blood cells were directed into channels 3 and 5 and the largest of the white cells were released and moved along channels 2 and 4. These cells appeared mainly to be neutrophils, and moved along channels 2 and 4 at a speed of the order 15 microns per second. On reducing the frequency of the reverse TWD signal from 400 kHz down to 150 kHz, the smaller white blood cells were released from the electrodes and travelled along channels 2 and 4. This cell separation process for dilute blood has been repeated for different levels of blood dilution and suspending medium composition, and it can be appreciated that in each case the specific frequency and voltage values cited above for the superimposed DEP and TWD signals were adjusted to achieve the results described above.

☐ 2. Document ID: US 6811753 B2

L13: Entry 2 of 24 File: USPT Nov 2, 2004

US-PAT-NO: 6811753
DOCUMENT-IDENTIFIER: US 6811753 B2

TITLE: Blood testing tool

L13: Entry 2 of 24 File: USPT Nov 2, 2004

DOCUMENT-IDENTIFIER: US 6811753 B2
TITLE: Blood testing tool

Detailed Description Text (8):
In order to maintain the stability of components in blood plasma or blood serum to be developed and retained, as described above, the development portion 12 may contain a stabilizing agent, for example, saccharides such as sucrose, trehalose, lactose, glucose, or the like, salts such as sodium chloride, potassium chloride, or the like, buffers such as glycine, a phosphate buffer, a citrate buffer, or a Good's buffer, or the like. The content of the stabilizing agent can be determined suitably according to its kind or the like, but is, for instance, in the range of 0.01 to 100 mg per cubic centimeter of the development portion 12. One kind of stabilizing agent may be used, or two or more kinds may be used together. The stabilizing agent may be provided not only in the development portion 12 but also the whole porous membrane.

Full | Title | Citation | Front | Review | Classification | Date | Reference | Sequences | Attachments | Claims | KWIC | Draw De

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☐ 3. Document ID: US 6677356 B1

L13: Entry 3 of 24 File: USPT Jan 13, 2004

US-PAT-NO: 6677356
DOCUMENT-IDENTIFIER: US 6677356 B1
** See image for Certificate of Correction **

TITLE: Treatment of cardiovascular and related pathologies

L13: Entry 3 of 24 File: USPT Jan 13, 2004

DOCUMENT-IDENTIFIER: US 6677356 B1
** See image for Certificate of Correction **
TITLE: Treatment of cardiovascular and related pathologies

Detailed Description Text (137):
It has been well demonstrated by various investigators that feeding 8-10% sucrose in water induces hypertension in rats. Zein et al., Am. Coll. Nutr., 17 (1), 36-37, 1998; Hulman et al., Pediatr. Res., 36:95-101; Reaven et al., Am. J. Hypertens; 1991:610-614. In applying this model, the concurrent administration of pyridoxal-5'-phosphate and captopril or verapamil significantly decreases the sucrose-induced increase in systolic blood pressure (SBP).

Detailed Description Text (139):
The effect of concurrent administration of pyridoxal-5'-phosphate and captopril or verapamil on systolic blood pressure (marker of hypertension) in 10% sucrose induced hypertension in rats is determined.

☐ 4. Document ID: US 6464976 B1

L13: Entry 4 of 24 File: USPT Oct 15, 2002

US-PAT-NO: 6464976
DOCUMENT-IDENTIFIER: US 6464976 B1

TITLE: Methods and compositions for reducing immune response

L13: Entry 4 of 24 File: USPT Oct 15, 2002

DOCUMENT-IDENTIFIER: US 6464976 B1
TITLE: Methods and compositions for reducing immune response

Detailed Description Text (35):
Five mice were each injected intraperitoneally with 40 .mu.g of anti-hAd antibodies eluted from the rAD-protein column, 80 .mu.g of anti-hAd antibodies eluted from rAd-protein column, column-depleted serum, untreated serum from primed animals and serum from vPBS (phosphate buffered saline further containing 2 mM MgCl.sub.2, 3% sucrose) injected animals and a sample of blood obtained one hour following passive immunization (to use as a standard) and allowed to rest overnight. The following day (approximately 12 hours post passive immunization), the mice were injected via the tail vein with 5.times.10.sup.10 particles of the BGCG recombinant human adenovirus. Serum was collected at 2 hours post injection and the serum was analyzed for the presence of serum neutralizing anti-adenoviral antibodies. The mice were sacrificed at 3 days following injection of BGCG and the serum analyzed for the presence of serum neutralizing anti-adenoviral antibodies and liver tissue was analyzed for B-gal expression. The results of these experiments can be seen in FIG. 4 of the attached drawings. Animals receiving passive immunization with unpurified serum or 40 .mu.g or 80 .mu.g of anti-hAd antibodies eluted from the column demonstrated a high titer of serum neutralizing anti-hAd antibodies one hour following IP injection. However, those animals which were passively immunized with column-depleted serum, had a no detectable serum neutralizing antibodies at one hour post injection. The presence of neutralizing antibodies in all five groups of mice on the third day following BGCG injection is due to a primary humoral response to BGCG by the. Consequently, the level of serum anti-hAd neutralizing antibodies in those animals passively immunized with purified serum was substantially diminished relative to those animals receiving unpurified serum indicating that removal of antiviral antibodies is useful in vivo to minimize the pre-existing immune response to such viruses. The serum purified over a SAVID column showed a improved reduction in serum neutralizing anti-hAd antibodies relative to the Protein A and KappaLock.TM.-Sepharose columns in vivo. Also evident from the data in FIG. 4 was that the intravenous injection of BGCG resulted in a further decrease in neutralizing antibodies in the sera (see undepleted serum two hour post virus). Moreover, in the group that received rAd antibody eluted from columns, virtually all the neutralizing antibody was depleted from the serum following injection of the BGCG by intravenous route. Thus systemic administration of rAd depletes serum

neutralizing antibody and suggests that high affinity antibodies are depleted very efficiently. Thus readministration one hour or one day after injection of rAd may further promote redosing.

☐ 5. Document ID: US 6184214 B1

L13: Entry 5 of 24 File: USPT Feb 6, 2001

US-PAT-NO: 6184214
DOCUMENT-IDENTIFIER: US 6184214 B1
** See image for Certificate of Correction **

TITLE: Pharmaceutical formulations

L13: Entry 5 of 24 File: USPT Feb 6, 2001

DOCUMENT-IDENTIFIER: US 6184214 B1
** See image for Certificate of Correction **
TITLE: Pharmaceutical formulations

Brief Summary Text (52):
A formulation used for rejuvenating red blood cells may, in addition to a compound of the invention, contain one or more of sodium or magnesium L-ascorbate phosphate, maltose, mannitol or sucrose, adenine, trisodium citrate, and sodium chloride (to adjust the osmolarity).

☐ 6. Document ID: US 6087087 A

L13: Entry 6 of 24 File: USPT Jul 11, 2000

US-PAT-NO: 6087087
DOCUMENT-IDENTIFIER: US 6087087 A

TITLE: Treatment of hemoglobin with nitric oxide

L13: Entry 6 of 24 File: USPT Jul 11, 2000

DOCUMENT-IDENTIFIER: US 6087087 A
TITLE: Treatment of hemoglobin with nitric oxide

Detailed Description Text (84):
Heparinated blood (10 ml), which is obtained from the Red Cross Blood Bank, is suspended in a 2-fold volume of chilled isotonic sucrose solution (the isotonic sucrose solution consists of 250 mM sucrose, 5 mM KCl, 2 mM NaH.sub.2 PO4, 1 mM MgCl.sub.2 6H2O, and 10 mM glucose.), centrifuged at 1,500 g for 10 minutes at 4 degrees C. After carefully decanting the supernatant and the buffy layer of

leukocytes, the loosely packed precipitate of erythrocytes is re-suspended in a fresh isotonic sucrose solution. The centrifugal washing procedure is repeated two more times.

☐ 7. Document ID: US 6077546 A

L13: Entry 7 of 24 File: USPT Jun 20, 2000

US-PAT-NO: 6077546
DOCUMENT-IDENTIFIER: US 6077546 A
**** See image for Certificate of Correction ****

TITLE: Quick-fermented feed and method of preparing

L13: Entry 7 of 24 File: USPT Jun 20, 2000

DOCUMENT-IDENTIFIER: US 6077546 A
**** See image for Certificate of Correction ****
TITLE: Quick-fermented feed and method of preparing

Brief Summary Text (72):
curd, corn gluten meal, lees of fermented corn, rapeseed meal, sesame meal, peanut meal, cotton seed meal, and sunflower meal are suitably used. If necessary, the present quick-fermented feeds can be prepared from a combination of the alkali-treated wastes of agricultural products, plant concentrate materials, and one or more materials consisting of by-products in the milk product industry such as skim milk powder, milk casein, and whey; animal concentrate materials such as fish meal, blood meal, and feather meal; saccharides such as molasses, sugar, glucose, maltose, and lactose; minerals such as bone meal, calcium salts, magnesium salts, sodium salts, phosphates, iron salts, copper salts, zinc salts, and cobalt salts; and vitamins such as water-soluble and lipid-soluble vitamins.

Full | Title | Citation | Front | Review | Classification | Date | Reference | Sequences | Attachments | Claims | KWIC | Draw D

☐ 8. Document ID: US 5897860 A

L13: Entry 8 of 24 File: USPT Apr 27, 1999

US-PAT-NO: 5897860
DOCUMENT-IDENTIFIER: US 5897860 A

TITLE: Method for controlling bleeding and microbial infections by administering thrombin, casein kinase II, and sphingosine

L13: Entry 8 of 24 File: USPT Apr 27, 1999

DOCUMENT-IDENTIFIER: US 5897860 A
TITLE: Method for controlling bleeding and microbial infections by administering thrombin, casein kinase II, and sphingosine

Detailed Description Text (49):
To 10 ml of human blood was added an equal amount of phosphate buffered saline (pH 7.4) containing 154 mM NaCl, and the resulting mixture was subjected to sucrose density gradient (1.077 g/l) centrifugation at 1,800 rpm, 20.degree. C. for 20 minutes to separate lymphocytes. The lymphocytes were mixed with 50 ml of the same buffer and subjected to centrifugation (1,600 rpm) at 4.degree. C. for 5 minutes to remove impurities. To 100 .mu.l of lymphocyte solution (3.times.10.sup.5 cells/ml) were added 50 .mu.l of casein kinase II solution (10 mM, 1 mM or 0.1 mM) and 50 .mu.l of .sup.3 H-thymidine. The resulting mixture was allowed to stand at room temperature for 3 days, and the radioactivity was measured every day. For comparison, a mixture of concanavalin A ("Con A") (10 .mu.g/ml) and interleukin-2 ("IL-2") (50 IU/ml) was used instead of casein kinase II (CK-II) solution.

Full | Title | Citation | Front | Review | Classification | Date | Reference | Sequences | Attachments | Claims | KWIC | Draw D

☐ 9. Document ID: US 5464634 A

L13: Entry 9 of 24 File: USPT Nov 7, 1995

US-PAT-NO: 5464634
DOCUMENT-IDENTIFIER: US 5464634 A

TITLE: Red blood cell surrogate

L13: Entry 9 of 24 File: USPT Nov 7, 1995

DOCUMENT-IDENTIFIER: US 5464634 A
TITLE: Red blood cell surrogate

Detailed Description Text (6):
In this example, red blood cell surrogates were made in the same manner as Example 1 except that pyridoxal-5-phosphate is substituted for cellobiose as the oxygen carrier anchor coating.

Full | Title | Citation | Front | Review | Classification | Date | Reference | Sequences | Attachments | Claims | KWIC | Draw D

☐ 10. Document ID: US 5462751 A

L13: Entry 10 of 24 File: USPT Oct 31, 1995

US-PAT-NO: 5462751
DOCUMENT-IDENTIFIER: US 5462751 A

TITLE: Biological and pharmaceutical agents having a nanomeric biodegradable core

L13: Entry 10 of 24 File: USPT Oct 31, 1995

DOCUMENT-IDENTIFIER: US 5462751 A
TITLE: Biological and pharmaceutical agents having a nanomeric biodegradable core

Detailed Description Text (126):
In this example, red blood cell surrogates are made in the same manner as Example 14 except that pyridoxal-5-phosphate is substituted for cellobiose as the oxygen carrier anchor coating.

Term	Documents
BLOOD	190807
BLOODS	506
(8 NEAR15 9 NEAR15 BLOOD).USPT.	24
((L8 NEAR15 L9 NEAR15 BLOOD)).USPT.	24